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**Research Article** 

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# Adsorption Behavior of Acid Red 97 Dye on Canola Stalks

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**Abstract** In this investigation, the mechanism of the adsorption of Acid Red 97 dye (AR97) from aqueous solution using an Canola stalks was studied. Different parameters affecting dye removal were studied. These parameters include contact time, initial dye concentration, adsorbent dose, stirring rate and adsorbent particle size. Langmuir and Freundlich isotherm models were applied to the equilibrium data. Equilibrium data fitted well with the Langmuir model and adsorption capacity (qmax) obtained from the Langmuir isotherm was 19.8 mg/g at initial pH 7. The optimum conditions for the removal of BPA within the experiment range of variables studies were 10 mg/L of initial dye concentration, 3.5 g/L of adsorbent dose, stirring rate 180 rpm, adsorbent particle size 20 mesh and 75 min of contact time. Under these conditions the maximum removal efficiency was 96.4%.

Keywords Acid Red 97 dye, Adsorption, Canola stalks, Isotherms

## Introduction

Common pollutants in textile effluents are dyes which are toxic, non- degradable due to their complex structure [1, 2]. As a result, the toxic effluents pose a huge risk to the environment if discharged untreated [3, 4]. Dyes are synthetic aromatic water soluble dispersible organic colorants, having potential application in various industries [5, 6]. The dyestuff usage has been increased day by day due to tremendous increase of industrialization and human desire for colour [7, 8]. It is estimated that there are currently more than 10,000 commercially available dyes with an annual production of over 0.7 million tons worldwide, of which 10 to 15% is lost in industrial effluents during manufacture and processing operations [9, 10]. The presence of very small amounts of dye in water (<1ppm for some dyes) causes aesthetic deterioration and diminishes the solubility of dissolved oxygen, water transparency, and sunlight permeability, affecting aquatic life and the food chain [11, 12].

There are several methods to remove dyes such as physical and chemical processes to treat wastewaters including organic pollutant and dyes [13, 14]. The most commonly used methods for color removal are biological oxidation and chemical precipitation [15, 16]. However, these processes are effective and economic only in the case where the solute concentrations are relatively high [17, 18]. The adsorption technique has proven to be an effective and attractive process for the treatment of these dye-bearing wastewaters [19, 20]. The adsorption characteristics of dyes on various adsorbents have been extensively investigated for many purposes involving separation and purification [21, 22]. Adsorption potential of selected activated carbons were tested and found that dye was effectively removed from aqueous solution [23]. However, the problem associated with the activated carbon decrease the attraction of its application [24]. Therefore, the researchers are trying to find a low-cost and effective adsorbent. Recently, the various natural material such as Red mud, Banana peel, Apple residues, Orange peel, Azolla and etc have been used as effective adsorbents to remove the dyes [25-27].

The canola stalk is one of lignocellulosic wastes that are widely produced in Iran and all of the world due to the growth of the production and consumption of vegetable oils; therefore, the Canola stalk is easily available and due to its characteristics has been used in several studies to remove the pollutants [28, 29]. The aim of the study was: (1) investigation of Canola ability to remove the AR97 dye from aqueous solution, (2) determination of adsorption behavior of AR97, (3) determination of impact of stirring rate, contact time, initial dye concentration, adsorbent particle size and biomass dosage on sorption.

#### **Materials and Methods**

In this study, the Canola was used as low cost natural or agricultural wastes for AR97 dye removal from aqueous solutions. The stalks of Canola were collected from research farm of Tabriz (Iran) agricultural school. Canola stalks were washed with distilled water and dried in a drier for 2 h at 105 °C until a constant weight was reached. It was then ground in a ball mill and sieved to particle sizes ranging from 10 to 100 mesh. The biomass were then treated with 0.5 M HCL for 2 h followed by the washing with distilled water and then was oven dried at 105°C for 3 h.

The AR97 dye was purchased from Alvan sabet Company (Iran). The stock solution of AR97 was prepared by dissolving 1 g of dye in 1000 ml of distilled water. The experimental solution say (10 to 200 mg/L) from stock solution by diluting to desired concentrations in accurate proportion. The chemical structure and some properties of the AR97 dye are presented in Table 1 and fig 1.

<b>TADLE 1:</b> UNALACIENSICS OF ACID RED 97
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Molecular Formula	CAS Number	<b>Colour Index Number</b>	$\lambda_{\max}$ (nm)	Molecular Weight					
$C_{32}H_{20}N_4Na_2O_8S_2$	10169-02-5	22890	630 nm	698.63					



Figure 1: Structure of Acid Red 97

#### **Adsorption studies**

Batch adsorption experiments were carried out at room temperature  $(27^{\circ}C\pm1)$ . Exactly 100 ml of dye solution of known initial concentration (50 mg/L) was stirred at a constant stirring rate (200 rpm) with a required dose of adsorbents (0.5-5 g/L) for a specific period of contact time (5–120 min) using a magnetic stirrer, at fixed pH=7. The pH of the solutions were adjusted to the required value by adding either 0.1 M HCl or 0.1 M NaOH solution. After equilibrium, the final concentration (C<sub>e</sub>) were measured using absorbance values with a spectrophotometer (UV-Vis-DR 5000) and compared with the absorbance values of the initial solutions. The absorbance was measured at 630 nm. Experiments were carried out in duplicate and mean values are presented. The percentage removal of dye were calculated using the following relationship [30]:

$$R = \frac{(Co - Ce)}{Co} \times 100$$

Where  $C_0$  and  $C_e$  are the initial and final (equilibrium) concentrations of dye (mg/L), respectively. Blanks containing second distilled water were used for each series of experiments as controls. The amount of dye adsorption per unit mass of Canola stalks at equilibrium, qe (mg/g) was calculated by the following equation [31]:

$$q_e = \frac{(Co - Ce) \times V}{V}$$

Where  $C_0^{r_1}$  and  $C_e$  are the initial and equilibrium dye concentrations in solution, respectively (mg/L), V the volume of the solution (L) and m is the mass (g) of the adsorbent used.

#### **Results and Discussion**

**Effect of contact time and initial dye concentration:** The effect of contact time on the percentage removal of the dye was investigated at initial dye concentration 50 mg/l as shown in Fig. 2. The percentage removal of dye by different Canola stalks was rapid in the beginning due to larger surface area available of adsorbent but it gradually decreased with time until it reached equilibrium. The plots reveal that maximum percent removal of the dye after about 1 h of continuous stirring [13, 32]. After adsorption, the rate of dye uptake is controlled by the rate of dye transported from the exterior to the interior sites of the adsorbent particles.

Fig. 2. shows the effect of contact time on the removal of disperse dye by Canola stalk for different initial dye concentrations. The adsorption amount increased with an increase in initial dye concentration. This is because a higher initial concentration enhanced the driving force between the aqueous and solid phases and increased the number of collisions between dye ions and adsorbents [33].





*Figure 2: Effect of contact time and initial dye concentration (Stirring rate=200 rpm, pH=7, Temp: 27 °C, adsorbent size =10 mesh and adsorbent dose =3.5 g/L)* 

**Effect of Stirring rate:** The effect of agitation speed on the dye adsorption (Fig. 3) at the adsorbent dosage of 3.5 g/L, initial dye concentration of 50 mg/L, pH 7. This effect can be attributed to the increase in turbulence and the decrease in boundary layer thickness around the adsorbent particles as a result of increase in the degree of mixing. This result also indicates that external mass transfer was the rate limiting step [34].



Figure 3: Effect of Stirring rate ( $C_0 = 50 \text{ mg/L}$ , pH=7, Contact time=75 min and Temp: 27 °C, adsorbent size =10 mesh and adsorbent dose =3.5 g/L)

**Effect of adsorbent dose:** The influence of adsorbent dosage on dye removal by Canola stalk in individual dyes is presented in Fig. 4. The percentage of removal of the dye increased from 32.4% to 89.2% as the Canola stalk dose was increased from 0.5 to 5 g/L. The increased removal at high dosages is expected, because of the increased adsorbent surface area and availability of more adsorption sites [35].



Figure 4: Effect of adsorbent dose ( $C_0 = 50 \text{ mg/L}$ , pH=7, Contact time=75 min and Temp: 27 °C, adsorbent size =10 mesh and Stirring rate=200 rpm)

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**Effect of adsorbent particle size:** The effect of different adsorbent particle sizes on percentage removal of dye is investigated and showed in Fig 5. It reveals that the adsorption of dye on tea powder decrease from 96.4 to 66.9% on coffee powder with the increased particle size from 10 mesh to 100 mesh at an initial concentration of 50 mg/L. The smallest size obtained was 10 mesh due to the limitation of available grinder configuration. It is well known that decreasing the average particle size of the adsorbent increases the surface area, which in turn increases the adsorption capacity [36].



*Figure 5: Effect of adsorbent particle size* ( $C_0 = 50 \text{ mg/L}$ , pH=7, *Contact time=75 min and Temp: 27 °C*, *Stirring rate=200 rpm and adsorbent dose =3.5 g/L*)

### **Adsorption isotherms**

The adsorption data were analyzed using adsorption isotherm models, Langmuir and Freundlich.

The Langmuir model is based on the assumption that maximum adsorption corresponds to a saturated monolayer of solute molecules on the adsorbent surface. The expression of the Langmuir model is given by the following equation [37, 38]:

$$q_e = \frac{q_{\max b c_e}}{1+hC}$$

Where  $q_e (mg/g)$  and  $C_e (mg/L)$  are the amount of adsorbed dye per unit mass of sorbent and dye concentration in solution at equilibrium, respectively.  $q_{max}$  is the maximum amount of the adsorbed dye per unit mass of sorbent to form a complete monolwar on the surface bound at high  $C_e (mg/g)$  and h (L/mg) is a

mass of sorbent to form a complete monolayer on the surface bound at high  $C_e$  (mg/g), and b (L/mg) is a constant related to the affinity of the binding sites on the adsorbent surface.

The linear form of the Langmuir equation is written as follows [39,40]:

 $\frac{Ce}{qe} = \frac{1}{q_{max} b} + \frac{Ce}{q_{m}}$ 

A plot of  $(C_e/q_e)$  versus  $C_e$  should be a straight line with a slope of  $1/q_{max}$  and intercept  $1/q_{max}$  b (fig 6). The essential characteristics of the Langmuir isotherm can be expressed in terms of a dimensionless constant

separation factor  $R_L$  that is given by the following equation [41,42]:

 $R_L = \frac{1}{1 + bC_0}$ 

Where  $C_0$  represents the initial concentration (mg/L) and b the Langmuir constant related to adsorption energy (l/mg).  $R_L$  value implies the shape of the isotherms to be either unfavorable ( $R_L$ ), linear ( $R_L$ =1), favorable ( $O < R_L < 1$ ) or irreversible ( $R_L$ =0). As can be seen in Table 2, for the sorption system,  $R_L$  values at different temperatures are between 0 and 1, showing favorable adsorption ( $C_0$ =50 mg/L). All  $R_L$  values at different temperatures and concentrations were between 0 and 1.

The Freundlich model assumes heterogeneous adsorption due to the diversity of active sites on the surface. The

Freundlich equation is expressed as [43,44]:

$$q_e = K_F C_e^{1/2}$$

Where  $K_F$  (mg/g) is an indicator of the biosorption capacity and 1/n (l/mg) is the biosorption intensity. A value for 1/n below one indicates a normal Frendlich isotherm while 1/n above one is an indicative of cooperative adsorption.

The linear form of the Freundlich equation is written as follows [45,46]:

 $\text{Log } q_e = \frac{1}{n} \log Ce + \log K_F$ 

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Where the values of  $K_F$  and 1/n are determined from the intercept and slope of the linear regressions (Table 2). As seen, a very high regression correlation coefficient was shown by the Langmuir model. This indicates that the Langmuir model was very suitable for describing the sorption of disperse AR97 dye on Canola stalks. The maximum capacity  $q_{max}$  determined from the Langmuir isotherm defines the total capacity of the Canola stalks for the dye as 19.8 mg/g sorbent. The fact that the Langmuir isotherm fits the experimental data compared to Freundlich isotherm may be due to the homogeneous distribution of active sites on the surface of sorbent.



 Table 2: Isotherms constants for the removal AR97 on Canola stalks

Langmuir model				Freundlich model		
$\mathbf{q}_{\mathbf{m}}$	R <sub>L</sub>	b	$\mathbf{R}^2$	n	K <sub>F</sub>	$\mathbf{R}^2$
19.8	0.594	0.028	0.992	3.44	1.086	0.934

### Conclusion

The results of different experiments showed that Canola stalks has an ability to adsorb disperse AR97 dye from aqueous solutions. Different variables, such as contact time, adsorbent dose, adsorbent particle size, initial dye concentration and stirring rate influenced the adsorptive quantity. The sorption process was found to be adsorbent particle size and stirring rate dependent. The adsorption process has nearly reached equilibrium in 75 min. The experimental data of adsorbing disperse AR97 dye are fit well to Langmuir isotherm model more than Freunlich model and the maximum adsorptive quantity of Canola stalks was 19.8 mg/g according to Langmuir model.

#### References

- Khaled A, Nem AE, El-Sikaily A, Abdelwahab O. Removal of Direct N Blue- 106 from artificial textile dye effluent using activated carbon from orange peel: adsorption isotherm and kinetic studies. Journal of Hazardous Materials. 2009: 165;100–110.
- [2]. Tor A, Cengeloglu Y. Removal of congo red from aqueous solution by adsorption onto acid activated red mud. Journal of Hazardous Materials. 2006;138(2):409-15.
- [3]. Zazouli MA, Balarak D, Mahdavi Y, Karimnejad F.The application of Azolla filiculoides biomass in acid blue 15 dye (AB15) removal from aqueous solutions. J Bas Res Med Sci 2014; 1(1):29-37.
- [4]. Mezohegyi G, Van der Zee FP, Font J, Fortuny A, Fabregat A. Towards advanced aqueous dye removal processes: A short review on the versatile role of activated carbon.Journal of Environmental Management. 2012.102; 148-164.
- [5]. Inbaraj BS, Chiu CP, Hob GH, Yang J, Chen BH. Removal of cationic dyes from aqueous solution using an anionic poly--glutamic acid-based adsorbent. Journal of Hazardous Materials 2006. B137; 226–234.
- [6]. Balarak D, Joghataei A, Azadi NA, Sadeghi S. Biosorption of Acid Blue 225 from Aqueous Solution by Azolla filiculoides: Kinetic and Equilibrium Studies. American Chemical Science Journal. 2016; 12(2):1-10.
- [7]. Kamal Amin N. Removal of reactive dye from aqueous solutions by adsorption onto activated carbons prepared from sugarcane bagasse pith. Desalination. 2008. 223; 152–161.



- [8]. Doulati Ardejani F, Badii KH, Yousefi Limaee N, Shafaei SZ, Mirhabibi AR. Adsorption of Direct Red 80 dye from aqueous solution onto almond shells: Effect of pH, initial concentration and shell type. Journal of Hazardous Materials 2008 151;730–737.
- [9]. Ozcan A, Oncu EM, Ozcan AS. Kinetics, isotherm and thermodynamic studies of adsorption of Acid Blue193 from aqueous solutions onto natural sepiolite. Colloids and Surfaces. 2006; A277, 90–97.
- [10]. Zazouli M, Balarak D, Mahdavi Y, Karimnejad F. Application of Azolla Filiculoides biomass for Acid blue 15 dye (AB15) Removal from aqueous solutions. JBRMS. 2014; 1 (1):29-37.
- [11]. Wu C. Adsorption of reactive dye onto carbon nanotubes: equilibrium, kinetics and thermodynamics. Journal of Hazardous Materials. 2007;144, 93–100.
- [12]. Mane VS, Mall ID. Kinetic and equilibrium isotherm studies for the adsorptive removal of Brilliant Green dye from aqueous solution by rice husk ash, J. Environ. Manage.2007: 84 ;390–400.
- [13]. Ozkan Al, Handan U. Equilibrium, kinetic and thermodynamic studies of the biosorption of textile dye reactive red 195 into pinus sylvestris. Journal of Hazardous Materials. 2010; 181: 666-72.
- [14]. Zazouli MA, Yazdani J, Balarak D, Ebrahimi M, Mahdavi Y. Removal Acid Blue 113 from Aqueous Solution by Canola. Journal of Mazandaran University Medical Science.2013:23(2);73-81.
- [15]. Dogan M, Abak H, Alkan M. Biosorption of Methylene Blue from Aqueous Solutions by Hazelnut Shells: Equilibrium, Parameters and Isotherms. Water Air Soil Poll. 2008; 192(1-4): 141-53.
- [16]. Padmesh TVN, Vijayaraghavan K, Sekaran G, Velan M. Application of Azollarongpong on biosorption of acid red 88, acid green 3, acid orange 7 from synthetic solutions. Chem Engin J. 2006; 11(122):55-63.
- [17]. Ponnusami V, Krithika V, Madhuram R, Srivastava SN. Biosorption of reactive dye using acid-treated rice husk: Factorial design analysis. Journal of Hazardous Materials. 2007; 8(142):397–403.
- [18]. Tan C-y, Li G, Lu X-Q, Chen Z-I. Biosorption of Basic Orange using dried A. filiculoides. Ecol Engin. 2010; 5(36):1333–40.
- [19]. Zazouli MA, Balarak D, Mahdavi Y. Effect of Azolla filiculoides on removal of reactive red 198 in aqueous solution. J Adv Environ Health Res. 2013; 1(1):1-7.
- [20]. Safa Y, Bhatti HN. Adsorptive removal of direct textile dyes by low cost agricultural waste: Application of factorial design analysis. Chem Engin J. 2011; 12(167):35–41.
- [21]. Karadag D, Turan M, Akgul E, Tok S, Faki A. Adsorption equilibrium and kinetics of Reactive Black 5 and reactive red 239 in aqueous solution onto surfactant-modified zeolite. J. Chem. Eng. 2007; 52;1615–1620.
- [22]. Iscen CF, Kiran I, Ilhan S. Biosorption of reactive black 5 dye by penicillium restrictum: The kinetic study. Journal of Hazardous Materials.2007; 143;335–340.
- [23]. Aydin HA, Yavuz O. Removal of acid red 183 from aqueous solution using clay and activated carbon, Indian. J. Chem. Technol. 2004;11;89–94.
- [24]. Mishra AK, Arockiadoss T,Ramaprabhu S. Study of removal of azo dye by functionalized multiwalled carbon nanotubes, Chem. Eng. J. 2010; 162;1026–1034.
- [25]. Mohammadi N, Khani H, Kumar Gupta V, Amereh E, Agarwal Sh. Adsorption process of methyl orange dye onto mesoporous carbon material-kinetic and thermodynamic studies. Journal of Colloid and Interface Science 2011 362; 457–462.
- [26]. Balarak D, Pirdadeh F, Mahdavi Y. Biosorption of Acid Red 88 dyes using dried Lemna minor biomass. Journal of Science, Technology & Environment Informatics 2015, 01(02), 81–90.
- [27]. Diyanati RA, Balarak D.Ghasemi SM. Survey of efficiency agricultural weast in removal of acid orang 7(AO7) dyes from aqueous solution: kinetic and equilibrium study: Iranian journal of health sciences. 2013;2(1):35-40.
- [28]. Balarak D, Mahdavi Y, Gharibi F, Sadeghi Sh. Removal of hexavalent chromium from aqueous solution using canola biomass: Isotherms and kinetics studies. J Adv Environ Health Res.2014; 2(4);45-52.
- [29]. Diyanati RA, Yousefi Z, Cherati JY, Balarak D. Comparison of phenol adsorption rate by modified Canola and Azolla: An Adsorption Isotherm and Kinetics Study.Journal of Health & Development. 2014; 3(3);17-25.
- [30]. Zazouli MA, Mahvi AH, Mahdavi Y, Balarak D. Isothermic and kinetic modeling of fluoride removal from water by means of the natural biosorbents sorghum and canola. Fluoride. 2015;48(1):15-22.
- [31]. Balarak D, Mahdavi Y, Bazrafshan E, Mahvi AH. Kinetic, isotherms and thermodynamic modeling for adsorption of acid blue 92 from aqueous solution by modified azolla filicoloides. Fresenius Environmental Bulletin.2016;25(5); 1321-1330.
- [32]. Song Qu, Fei Huang S, Yu K. Magnetic removal of dyes from aqueous solution using multi-walled carbon nanotubes filled with Fe2O3 particles. Journal of Hazardous Materials. 2008; 160: 643-347.

- [33]. Vadivelan V, Kumar KV. Equilibrium, kinetics, mechanism, and process design for the sorption of methylene blue onto rice husk. Journal of Colloid and Interface Science., 2005; 286, 90–100.
- [34]. Arami M, Yousefi N, Mahmoodi NM. Evaluation of the adsorption kinetics and equilibrium for the potential removal of acid dyes using a biosorbent. Chem. Eng. J. 2008;139; 2-10.
- [35]. Bulut Y, Gozubenli N, Aydin H. Equilibrium and kinetics studies for adsorption of direct blue 71 from aqueous solution by wheat shells. Journal of Hazardous Materials. 2007;144; 300-307.
- [36]. Wong Y, Szeto Y, Cheung W, McKay G. Adsorption of acid dyes on chitosan equilibrium isotherm analyses. Process Biochem. 2004;39;693–702.
- [37]. Malakootian M, Balarak D, Mahdavi Y, Sadeghi SH, Amirmahani N. Removal of antibiotics from wastewater by azolla filiculoides: kinetic and equilibrium studies. International Journal of Analytical, Pharmaceutical and Biomedical Sciences. 2015;4(7);105-113.
- [38]. Mishra AK, Arockiadoss T, Ramaprabhu S.Study of removal of azo dye by functionalized multiwalled multi-walled carbon nanotubes. Chem. Eng. J. 2010; 162,1026-1034.
- [39]. Meziti C, Boukerroui A. Removal of a Basic Textile Dye from Aqueous Solution by Adsorption on Regenerated Clay. Procedia Engineering. 2012;33(0):303-12.
- [40]. Moussavi GR, Mahmoudi M. Removal of azo and anthraquinone reactive dyes from industrial wastewaters using MgO nanoparticles. Journal of Hazardous Materials 2009. 168; 806–81.
- [41]. Hoda N, Bayram E, Ayranci E. Kinetic and equilibrium studies on the removal of acid dyes from aqueous solutions by adsorption onto activated carbon cloth. Journal of Hazardous Materials 2006. B137; 344–351.
- [42]. Balarak D, Mahdavi Y, Azadi NA, Sadeghi SH. Isotherms and thermodynamic study on the biosorption of amoxicillin using canola. International Journal of Analytical, Pharmaceutical and Biomedical Sciences.2016;5(3);8-14.
- [43]. Zazouli MA, Balarak D, Karimnezhad F, Khosravi F. Removal of fluoride from aqueoussolution by using of adsorption onto modified Lemna minor: adsorption isotherm and kinetics study. Journal of Mazandaran University Medical Sciences 2014;23(109):208-17.
- [44]. Balarak D, Jaafari J, Hassani G, Mahdavi Y, Tyagi I, Agarwal S, Gupta VK. The use of low-cost adsorbent (Canola residues) for the adsorption of methylene blue from aqueous solution: Isotherm, kinetic and thermodynamic studies.Colloids and Interface Science Communications. Colloids and Interface Science Communications.2015; 7;16–19.
- [45]. Balarak D, Mahdavi Y, Bazrafshan E, Mahvi AH, Esfandyari Y. Adsorption of fluoride from aqueous solutions by carbon nanotubes: determination of equilibrium, kinetic and thermodynamic parameters. Flouride. 2016;49(1);35-42.
- [46]. Diyanati RA, Yousefi Z, Cherati JY, Balarak D. The ability of Azolla and lemna minor biomass for adsorption of phenol from aqueous solutions. J Mazandaran Uni Med Sci. 2013; 23(106).17-23.

