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Research Article

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DNA Barcoding of *Liriomyza sylvatica* Sehgal 1971 (Diptera: Agromyzidae): a new record for Türkiye

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Abstract In this study, DNA barcoding of *Liriomyza sylvatica* Sehgal 1971 (Diptera: Agromyzidae) was collected from a forest area in Aydın, Türkiye between June and September 2023. Adults were obtained by culturing with galleried leaves in the laboratory. After the morphological diagnosis of *L. sylvatica*, HCO/LCO universal primers were used to reproduce the COI gene region of the samples, and 614 bp COI regions were obtained. Sequencing of the samples was performed at BM Labosis using ABI 3730XL DNA sequencer (Applied Biosystems). The primer pair used in PCR was also used in sequencing. As a result of the sequencing, files sequenced with both primers were compared with BioEdit software to create a single file. The obtained sequence was compared with the samples registered in the gene bank (NCBI, Nucleotide BLAST) to determine the species. The nucleotide distribution of the amplified COI region and Neighbour Joining (NJ) tree were calculated in the MEGA 11 package program. The control of whether the obtained sequence was pseudogene was also performed in the Gen Bank BlastX module. In our study, the insects obtained in Aydın/Türkiye were determined to *be L. sylvatica*, and Genbank access records were obtained. *L. sylvatica* is a new record for the Agromyzidae fauna of Türkiye.

Keywords COI, DNA barcoding, *Liriomyza sylvatica*, Türkiye

1 Introduction

Their larvae can cause a loss of up to 98% of the leaf area by opening galleries between the two epidermises of the leaf and can also lead to decreases in the chlorophyll content of the plant [1]. Adults can cause also indirect harm through their egg-laying behavior and helping vectors by viruses [2]. There are approximately 2700 species of the Agromyzidae family in the world, belonging to 27 genera, whereas 776 species in Europe. Up to now, 208 species have been identified in Türkiye [3, 4, 5, 6]. The species identification of leafminer flies is made by using their external morphological features (color, hair distribution number, etc.) and aedeagus. The fact that they are very small in size and that the diagnostic features between species are very similar to each other, as well as the lack of sufficient experts in this field, necessitate turning to morphological techniques in diagnosis. The COI gene region can be used as an identification marker in studies conducted with insects as well as in most animal groups. It has many important advantages such as having extensive phylogenetic data and ease of operation when compared to other protein-coding genes [7]. For these reasons, this study aimed to perform molecular identification and determine phylogenetic distances of *Liriomyza sylvatica*, which is thought to be a new record for Türkiye, by gene barcoding.

2. Materials and Methods

Samples were collected by sweep netting in Aydın provinces, Türkiye (37°41′15′′N, 27°41′5′′E, 770m) between June and September 2023.



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Laboratory studies

Species identifications were made according to Spencer [8]. and Sehgal [9]. Polymerase Chain Reaction (PCR) was used to amplify the COI gene region and this study was carried out with a PCR master mix and universal COI primer pair (LCO1490 and HCO2198). The PCR reaction was prepared in a total volume of 25 μ L containing 2 μ L template DNA and 12.5 μ L PCR master mix. PCR master mix contained Tris-HCl pH 8.5, (NH4)2SO4, 4 mM MgCl2, 0.2% Tween 20, 0.4 mM each dNTP, Ampliqon Taq DNA polymerase, non-interacting red dye, and stabilizer. The PCR cycle was performed as indicated in Table 1.

Table 1: COI	gene region	PCR	conditions
Table 1. COI	gene region	ICI	contamons

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Stages of PCR		Temperature/Time	Cycle number
Initial denaturation		95 °C / 5 min	1
Steps of cycles	Denaturation	95 °C / 30 sec	35
	Primer annealing	55 °C / 40 sec	35
	Extension	72 °C / 40 sec	35
Final extension		72 °C / 5 min	1

Sequencing of the samples was performed at BM Labosis using ABI 3730XL DNA sequencer (Applied Biosystems). The primer pair used in PCR was also used in sequencing. As a result of the sequencing, files sequenced with both primers were compared with BioEdit [10] software to create a single file. The obtained sequence was compared with the samples registered in the gene bank (NCBI, Nucleotide BLAST) to determine the species. The nucleotide distribution of the amplified COI region was calculated in the MEGA 11 [11]. package program. The control of whether the obtained sequence was pseudogene was also performed in the Gen Bank BlastX module. The obtained sequence was registered in Gen Bank and under the accession codes PQ182227. The partial COI sequence obtained from the study sample was aligned with the sequences of the related species *Liriomyza septentrionalis* (MG120229, MF636182), *L. phryne* (OK065481), *L. sylvatica* (KM854006, MF636607), *L. brassicae* (OR038460), *L. bryoniae* (OP161993), *L. trifolii* (OR038586), *L. sativae* (OR038567) and *L. fricki* (MF635341) in the Bioedit program to create a Neighbour Joining (NJ) [12] tree in the MEGA 11 package program [11] based on Kimura 2- parameter distances, with 1000 replicates of bootstrapping [13]. *Phytomyza gymnostoma* (MN943089) was used as an outgroup to root the tree.

3. Results & Discussion

The sequences read with both primers were aligned by eye in the Bioedit program.

Score 1070 b	its(57		Expect 0.0	Identities 583/585	(99%)	Gaps 0/585(0	Strand %) Plus/I	
Query	30	AACTTTATA	TTTTATAT	CGGAGCT1	GAGCTGGAA	TAGTGGGAACT	TCTCTTAGAATTTT	89
Sbjct	1	AACTTTATA	TTTTATAT	rcggagcti	GAGCTGGAA	FAGTGGGAACT		60
Query	90						CAAATTTATAATGT	149
Sbjct	61						CAAATTTATAATGT	120
Query	150						CCTATTATAATTGG	209
Sbjct	121						CCTATTATAATTGG	180
Query	210					GTGCTCCAGAT	ATAGCCTTTCCTCG	269
Sbjct	181				TÄÄTÄTTÄG		ATAGCCTTTCCTCG	240
Query	270	AATAAATAA	TATAAGTT	TTTGACTAT	TACCTCCAG	CCTTACCTTA	CTTTTAATAAGTAG	329
Sbjct	241						CTTTTAATAAGTAG	300
Query	330						CTTTCTTCTATTAT	389
Sbjct	301					TTTACCCTCCA	CTTTCTTCTATTAT	360
Query	390						TTAGCTGGAGTCTC	449
Sbjct	361						TTAGCTGGAGTCTC	420
Query	450	TTCTATTTT	AGGAGCAG	TAAATTTT/	ATTACAACTA	TTATCAATATA	CGATCTACTGGAAT	509
Sbjct	421					TTATCAATATA	CGATCTACTGGAAT	480
Query	510					* TACTAATTACT	GCAGTATTATTATT	569
Sbjct	481						GCAGTATTATTATT	540
Query	570	ATTATCTCT			CTATTACTA		614	
Sbjct	541	ATTATCTCT			GCTATTACTA		585	

Figure 1: Similarity diagram with the sequence referenced as KM870016, which is one of the closest sequences to our sample sequence in the Genbank Nucleotide Blast model (* symbol indicates dissimilar bases).



The common sequence obtained is 614 base pairs long. In this sequence, the percentage equivalent of nucleotide frequencies was determined as T 40.1%, C 15.6%, A 28.2% and G 16.1%. The G+C ratio of the sequence is 31.76%. When compared with previously identified samples recorded in the BlastN module in the Genbank, the 6 *Liriomyza sylvatica* sequences (KM870016, KM865480, KM854006, MF636607, MF635288, and MF632746) with the highest similarity were at the top. The highest similarity was 99.66% with KM870016 and KM865480 sequences (Figure 1). Although the similarity with the MF632746 reference sequence was 99.80%, this sequence was not evaluated because it was short (510 bp) and one of the 2 base differences in our sample did not cover the query made with this sequence. As seen in the figure, there are 2 dissimilar bases (303: A>T and 547: C>T) between the sequence of our sample and the reference sequence, and there is no gap. Whether the sequence of our sample is a pseudogene was tested in the BlastX module of the Genbank.

BlastX comparison showed that the sequence we had was not a pseudogene, but a partial gene encoding a part of the cytochrome oxidase enzyme (Figures 2, 3).

Score		Expect	Method		Identities	Positives	Gaps	Frame
258 bit	s(659)	1e-83	Compositional matrix	adjust.	175/203(86%)	197/203(97%)	0/203(0%)	+1
Query	4		DIGTLYFIFGA*AGIVGT DIGTLYFIFGA AG+VGT					
Sbjct	1		DIGTLYFIFGAWAGMVGT					
Query	184		iiggfgN*LVPLILGAPD +IGGFGN LVPL+LGAPD					
Sbjct	61		MIGGFGNWLVPLMLGAPD					
Query	364		SIIAHGGASVDLAIFSLH S+IAHGGASVDLAIFSLH					
Sbjct	121	VYPPLS	SVIAHGGASVDLAIFSLH	LAGVSSI	LGAVNFITTVINM	RATGITFDRMPLF	VWS 180	
Query	544		llllslpvlaGAITILL	612				
Sbjct	181		LLLLSLPVLAGAITMLL	203				

Figure 2: BlastX mapping diagram of our sample sequence.

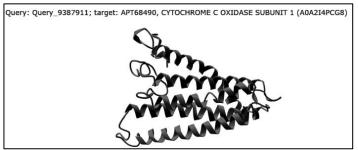


Figure 3: Partial cytochrome oxidase enzyme fragment encoded by the sequence.

In many previous studies on the COI gene region, the minimum distance between close species (sister group) within the genus in terms of the COI gene region was reported as greater than 2.5% [14, 15, 16, 17, 18, 19]. In this context, our sample was molecularly determined to be *Liriomyza sylvatica* in the light of the COI gene region. In our NJ tree, the genetic distance between the "*L. sylvatica*" pedigree and the "*L. septentrionalis*" and "*L. phyrne*" pedigrees in the sister group relationship is 9.6% and 7.9%. The genetic distance was 12.8% for "*L. trifoli*", 14.3% for "*L. brassicae*", 11.5% for "*L. sativa*" and 12.5% for "*L.fricki*".

Table 2: Estimates of Evolutionary Divergence over Sequence Pairs between Groups*

Species	[1]	[2]	[3]	[4]	[5]	[6]	[7]
L. septentrionalis [1]							_
L. phryne [2]	0.091						
L. sylvatica [3]	0.096	0.079					
L. trifolii [4]	0.145	0.135	0.128				
L. brassicae [5]	0.171	0.158	0.143	0.149			
L. sativae [6]	0.138	0.124	0.115	0.070	0.135		
L. fricki [7]	0.141	0.135	0.125	0.149	0.165	0.142	
L. bryoniae [8]	0.148	0.151	0.128	0.126	0.146	0.120	0.135

^{*}The number of base substitutions per site from averaging over all sequence pairs between groups is shown.



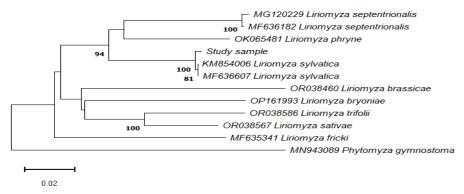


Figure 4: Phylogenetic analysis of samples

Studies on the use of COI gene region in the identification and phylogenetic analysis of species belonging to *Liriomyza* spp. have been increasing in recent years [20, 21]. This molecular method provides significant convenience, especially in the identification of species that are difficult to identify morphologically. With this study, *Liriomyza sylvatica* was recorded as a new record for Türkiye.

4. Conclusion

Due to reasons such as *Liriomyza* spp. being a small insect and the small number of experts working on this subject, DNA barcode studies can be used more effectively both for faster identification and to increase the reliability of the identification. In our study, the insects obtained in Aydın/Türkiye were determined to *be L. sylvatica*, and Genbank access records were obtained. *L. sylvatica* is a new record for the Agromyzidae fauna of Türkiye.

References

- [1]. Yıldırım, E. M., Ünay, A., & Civelek, H. S. (2010). The effect of Liriomyza trifolii (Burgess) (Diptera: Agromyzidae) on some leaf characteristics of bean (Phaseolus vulgaris L.). Journal of Food, Agriculture & Environment, 8 (3&4):839-841.
- [2]. Costa, A.S., Silva, D.M., & Duffus, J.E. (1988) Plant virus transmission by leafminer fly. Virology, 5(1): 145-149
- [3]. Spencer KA (1972) Agromyzidae from Southern Spain (Insecta: Diptera). Zoological Museum University of Copenhagen, 2(6): 91–104.
- [4]. Spencer KA (1976) The (Diptera: Agromyzidae) of Fennoscandia and Denmark. Fauna Entomologica Scandinavica, 5(1–2): 1–606.
- [5]. Spencer KA (1990) Host specialization in the world (Diptera: Agromyzidae). Kluver Academic Publishers, Netherland, 444 s.
- [6]. Dursun, C. (2015). Determination of the Biodiversity of Agromyzidae (Diptera) Family and their Parasitoid Species Belong to Eulophidae (Hymenoptera)Family in Muğla Province. Muğla Sıtkı Koçman University, Institute of Science, PhD Thesis,228p (in Turkish with English abstract).
- [7]. Polat, F., Dede, S., Bingöl, G., & Kekillioğlu, A. (2018). Kocaeli'de Yayılış Gösteren Bazı Böcek Türlerinin Mitokondrial Sitokrom Oksidaz Alt Ünite 1 Geni ile Filogenetik Analizi. Kocaeli Üniversitesi Fen Bilimleri Dergisi, 1(2), 62-66.
- [8]. Spencer, K.A. (1969) The Agromyzidae of Canada and Alaska. Memoirs of the Entomological Society of Canada, 64, 1–311. https://doi.org/10.4039/entm10164fv
- [9]. Sehgal, V.K. (1971). A Taxonomic Survey of the Agromyzidae (Diptera) of Alberta, Canada, with Observations on Host-Plant Relationships. Quaestiones entomologicae, 7: 291-405 1971
- [10]. Hall, T. A. (1999). BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. In Nucleic acids symposium series, 41 (41): 95-98.
- [11]. Tamura, K., Stecher, G., & Kumar, S. (2021). MEGA11: molecular evolutionary genetics analysis version 11. Molecular biology and evolution, 38(7), 3022-3027.



- [12]. Saitou, N., & Nei, M. (1987). The neighbor-joining method: a new method for reconstructing phylogenetic trees. Molecular biology and evolution, 4(4), 406-425.
- [13]. Felsenstein, J. (1985). Phylogenies from gene frequencies: a statistical problem. Systematic zoology, 34(3), 300-311.
- [14]. Chan, A., Chiang, L. P., Hapuarachchi, H. C., Tan, C. H., Pang, S. C., Lee, R., Lee, K.S., ChingNg, L., & Lam-Phua, S. G. (2014). DNA barcoding: complementing morphological identification of mosquito species in Singapore. Parasites & vectors, 7, 1-12.
- [15]. Klicka, J., & Zink, R. M. (1997). The importance of recent ice ages in speciation: a failed paradigm. Science, 277(5332), 1666-1669.
- [16]. Johns, G. C., & Avise, J. C. (1998). A comparative summary of genetic distances in the vertebrates from the mitochondrial cytochrome b gene. Molecular biology and evolution, 15(11), 1481-1490.
- [17]. Hebert, P. D., Penton, E. H., Burns, J. M., Janzen, D. H., & Hallwachs, W. (2004). Ten species in one: DNA barcoding reveals cryptic species in the neotropical skipper butterfly Astraptes fulgerator. Proceedings of the National Academy of Sciences, 101(41), 14812-14817.
- [18]. Wilson, J. J., Sing, K. W., Floyd, R. M., & Hebert, P. D. (2017). DNA barcodes and insect biodiversity. Insect biodiversity: science and society, 575-592.
- [19]. Liang, Y. X., Du, S. J., Zhong, Y. J., Wang, Q. J., Qiong, Z. H. O. U., Wan, F. H., Guo, J. Y., & Liu, W. X. (2023). Molecular phylogeny and identification of agromyzid leafminers in China, with a focus on the worldwide genus Liriomyza (Diptera: Agromyzidae). Journal of Integrative Agriculture, 22(10), 3115-3134.
- [20]. Carapelli, A., Soltani, A., Leo, C., Vitale, M., Amri, M., & Mediouni-Ben Jemâa, J. (2018). Cryptic diversity hidden within the leafminer genus Liriomyza (Diptera: Agromyzidae). Genes, 9(11), 554.
- [21]. Dayan, C., & Yıldırım, E. M. (2022). Liriomyza sativae Blanchard, 1938 (Diptera: Agromyzidae)'nin DNA Barkodlaması. Adnan Menderes Üniversitesi Ziraat Fakültesi Dergisi, 20(1), 167-172.